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INTERNATIONAL APPLICATION PUBLISHED UNDEI WO 9607432A1

(51) International Patent Classification 6 : A61K 47/48, 47/00

A1

(11) International Publication Number: WO 96/07432

(43) International Publication Date: 14 March 1996 (14.03.96)

(21) International Application Number: PCT/NO95/00149

(22) International Filing Date: 4 September 1995 (04.09.95)

(30) Priority Data:
943327 8 September 1994 (08.09.94) NO

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(81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LT, LU, LV, MD, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TT, UA, UG, US, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ, UG).

Published

With international search report.

Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(54) Title: TRANSFER OF MOLECULES INTO THE CYTOSOL OF CELLS

(57) Abstract

A method for releasing molecules into the cytosol of cells without killing the majority of the cells by allowing the molecules to be taken up in endosomes, lysosomes or other cell compartments and use light activation of photosensitizing compounds to rupture the membranes of the endosomes, lysosomes or other cell compartments, is described.

BNSDOCID: <WO___9607432A1_I_>

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BNSDOCID: <WO 9607432A1 >

Transfer of molecules into the cytosol of cells

The present invention relates to a method for introducing molecules in cells by disrupting endosomal and lysosomal membranes using photodynamic treatment, without killing the majority of the cells by the photodynamic treatment.

The majority of molecules do not readily penetrate cell membranes. Methods for introducing molecules into the cytosol of living cells are useful tools for manipulating and studying biological processes. Among the most commonly used methods today are microinjection, red blood cell ghost mediated fusion and liposome fusion, osmotic lysis of pinosomes, scrape loading, electroporation, calcium phosphate and virus mediated transfection. These techniques are useful for investigations of cells in culture, although in many cases impractical, time consuming, inefficient or they induce significant cell death. They are thus not optimal for use in biological and medical research or therapeutics in which the cells shall remain functional.

It is well known that porphyrins and many other photosensitizing compounds induce cytotoxic effects on cells and tissues. These effects are based upon the fact that the photosensitizing compound upon light exposure releases singlet $^1O_2$ which decomposes the membranes of the cells and cell structures and eventually kill the cells if the destruction is extensive. These effects have been utilized to treat several types of neoplastic diseases. The treatment is named photodynamic therapy (PDT) and is based on injection of a photosensitizing and tumorlocalizing dye followed by exposure of the tumor region to light. The cytotoxic effect is mediated mainly through the formation of singlet oxygen. This reactive intermediate has a very short lifetime in cells (<0.04μs). Thus, the primary cytotoxic effect of PDT is executed during light exposure and very close to the sites of formation of $^1O_2$. $^1O_2$ reacts with and oxidize proteins (histidine, tryptophan, methionine, cysteine, tyrosine), DNA (guanine), unsaturated fatty acids and cholesterol. One of the advantages of PDT is that tissues unexposed to light will not be affected. There is extensive documentation regarding use of PDT to destroy unwanted cell popuiation, for example neoplastic cells. Several patents relate to photodynamic compounds alone or conjugated with immunoglobulins directed to nepolastic cell receptor determinants making the complex more cell specific. Certain photochemical compounds, such as hematoporphyrin derivates have furthermore an inherent ability to concentrate in malign cells. These metods and compounds, which are directed to destroy the

unwanted cells are described in the Norwegian patent NO 173319, in Norwegian patent applications Nos 90 0731, 90 2634, 90 1018, 94 1548, 85 1897, 93 4837, 92 4151 and 89 1491.

In PCT/US93/00683 a drug delivery system is described which is comprised of an anticancer drug and a photoactivatable drug attached to copolymeric carriers. Upon administration this complex enters the cell interior by pinocytosis or phagocytosis and will be located inside the endosomes and lysosomes. In the lysosomes the bond between the antineoplastic compound and the polymer is hydrolyzed and the former can diffuse passively through the lysosome membrane into cytosol. Thus this method limits the method to small molecular compounds which are able to diffuse across the lysosome membranes. After allowing a time lag for diffusion a light source of appropriate wavelenght and energy is applied to activate the photoactivatable compound. The combined effect of the anticancer drug and photoactivatable drug destroy the cell. Thus all use of photoactivatable compounds known is directed to extensively destroy cell structures leading to cell death. It is not known of a method to release membrane unpermeable molecules into the cytosol after localized rupturing of endosomal/lysosomal membranes.

The object of the present invention is thus to provide a method to transport molecules into cytosol of living cells, in culture or in tissues, by exposing the cells to a photoactivatable compound, the molecule(s) which is (are) to be transported into the cytosol, both of which uptake may be facilitated by various carriers, exposing the cell to light of suitable wavelength and energi to disrupt the endosomal and lysosomal membranes and release the molecules into the cytosol without destroying the functionality of the majority of the cells. The photosensitizer and the molecule(s) which is (are) to be transported into the cytosol may be conjugated to or applied separately together with suitable carriers, optionally facilitating the uptake of the molecules of interest.

This object is obtained by the present invention characterized by the enclosed claims.

The present invention relates to a method for transporting any molecules into the cytosol of living cells after which the molecules shall be available in the cytosol and the cell shall maintain its functionability. This is performed by exposing the cell(s) to a photoactivatable compound which is taken up by the cell and will be

located in endosomes, lysosomes or other cellular compartments, conjugated to or separately together with carrier molecules, targeting immunoglobulins and the molecules to be transported into the cytosol and expose the cells to light of suitable wavelength to activate the photosentizing compound, such that only the endosomal, lysosomal or other cellular compartment membranes are ruptured and the molecules released in the cytosol without the cell loosing its functionability by the action of the photoactivated compound and possible action of the enosomal/lysosomal content.

In the following the present invention is described in detail and illustrated by the figures, of which;

Fig.1. represents illustration of how molecules can be introduced into the cellular cytosol by means of the present invention. The photosensitizer (S) and the molecules of choice (M) are endocytosed by the cells (I, illustrates the invagination of the plasma membrane initiating the endocytic process) and both substances end up in the same vesicles (II). When these vesicles are exposed to light, the membranes of the vesicles rupture and the contents are released (III);

Fig.2. illustrates protein synthesis in NHIK 3025 cells after treatment with gelonin in the absence or presence of $TPPS_{2a}$ and 50 sec light exposure. Symbols: ◯, $TPPS_{2a}$ + light; ●, - $TPPS_{2a}$ - light; ▽, + $TPPS_{2a}$ - light; ▼, - $TPPS_{2a}$ + light. The cells were treated with 3,2 µg/ml $TPPS_{2a}$ and the indicated concentration of gelonin overnight and in all cases given the same dose of light. Protein synthesis was measured by measuring incorporation of ^{3}[H]leucine into proteins, 24 h after light exposure;

Fig.3. shows dose-response curves for cells treated with $TPPS_{2a}$ and light only (◯) or in combination with 0.2 µg/ml (▽) or 2.0 µg/ml gelonin as described Fig.2; and

Fig.4. shows protein synthesis in NHIK 3025 cells after treatment with 3,2 µg/ml $TPPS_{2a}$ and light in the absence or presence of 0,2 µg/ml gelonin. Symbols: ●, $TPPS_{2a}$ - gelonin; ○, $TPPS_{2a}$ + gelonin. The cells were treated with $TPPS_{2a}$ in the absence or presence of gelonin

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overnight and exposed to the indicated doses of light. Protein synthesis was measured by measuring incorporation of ^{3}[H]leucine into proteins.

Fig. 5 illustrates protein synthesis in V79 cells after treatment with 25 µg/ml $AlPcS_2$ and light in the absence and presence of 1 µg/ml gelonin. Symbols: ● Photosensitizer + toxin; O photosensitizer,

Fig. 6 illustrates protein synthesis in H146 cells after treatment with 0,3 µg/ml $TPPS_{2a}$ and light in the absence and presence of 1 µg/ml gelonin. Symbols: as in Fig. 5,

Fig. 7 illustrates protein synthesis in V79 cells after treatment with 1 µg/ml $TPPS_{2a}$ and light in the absence and presence of 1 µg/ml gelonin. Symbols: as in Fig. 5,

Fig. 8 illustrates protein synthesis in NHIK3025 cells after treatment with 3,2 µg/ml $TPPS_{2a}$ and light in the absence and presence of 1 µg/ml agrostin. Symbols: as in Fig. 5,

Fig. 9 illustrates protein synthesis in NHIK3025 cells after treatment with 3,2 µg/ml $TPPS_{2a}$ and light in the absence and presence of 1 µg/ml saporin. Symbols: as in Fig. 5,

Fig. 10 illustrates protein synthesis in NHIK3025 cells after treatment with 0,25 µg/ml 3-THPP and light in the absence and presence of 1 µg/ml gelonin. Symbols: as in Fig. 5,

Fig. 11 illustrates protein synthesis in COS-7 cells after treatment with 3 µg/ml $TPPS_{2a}$ and light in the absence and presence of 1 µg/ml gelonin. Symbols: as in Fig. 5,

Fig. 12 illustrates protein synthesis in NHIK 3025 cells after treatment with gelonin in the absence or presence of TPPS4 and 50 sec light exposure. Symbols: ■ $TPPS_4$ + light; ▾ - $TPPS_4$ - light; ◊ + $RPPS_4$ - light. The cells were treated with 75 µg/ml $TPPS_4$ and the indicated concentration of gelonin overnight and in all cases given the same dose of light. Protein synthesis was measured by measuring

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incorporation of $^{3}[H]$leucine into proteins, 24 h after light exposure; and

Fig. 13 illustrates protein synthesis in OHS cells after treatment with 3 μg/ml $TPPS_{2a}$ for 18 hours followed by 4 hours in the absence of $TPPS_{2a}$ and in the absence or presence of 3 μg/ml gelonin before exposure to light. The cells were incubated for the same 4 hours in 50 μM chloroquine or 10 mM NH_4Cl to inhibit lysosomal protein degration.

It is well documented that a number of drugs, including di- and tetrasulfonated aluminium phthalocyanine, sulfonated tetraphenylporphines ($TPPS_n$), nile blue, chlorin e_6 derivatives, uroporphyrin I, phylloerythrin and possibly hematoporphyrin and methylene blue is located in endosomes and lysosomes of cells in culture. This is in most cases due to endocytic activity. The inventors have shown that light exposure of cells containing photosensitizers in their lysosomes leads to a permeabilization of the lysosomes and release of the photosensitizer. In some cases, e.g. $TPPS_{2a}$ and $TPPS_1$, substantial amounts of lysosomal enzyme activities have been found in the cytosol after PDT, indicating that lysosomal contents can be released into the cytosol without losing their activity. This effect of photosensitizing dyes can be used to release endocytosed molecules from endosomes and lysosomes in general according to the present investigation.

The introduction of molecules into the cellular cytoplasm is achieved by first exposing the cells or tissue to a photosensitizing dye, the molecule(s) which one wants to deliver into the cytosol of the cells together with, or not carrier molecules and immunoglobins, all of which should preferentially localize in endosomes and/or lysosomes. Secondly, the cells or tissue is exposed to light of suitable wavelengths and energies inducing a photodynamic reaction. This photodynamic reaction will lead to disruption of lysosomal and/or endosomal membranes and the contents of these vesicles will be released into the cytosol.

The principles of the present invention are illustrated in Fig.1. It is necessary that the photosensitizer and the molecule to be introduced into the cells are located in the same compartments. It should also be emphasized that externally added molecules may accumulate in intracellular compartments other than lysosomes and endosomes, e.g. Golgi apparatus and endoplasmic reticulum. In such cases, photosensitizing compounds located in the same compartments may

in combination with light be used for the same purposes provided that the combination of light dosis and photosensitizing compound does not destroy the functionality of the cells.

The present invention is based on our in vitro demonstration, that a photosensitizer, for example $TPPS_{2a}$, (tetraphenylporphine with 2 sulfonate groups on adjacent phenyl groups) in combination with light can induce release of functionally intact lysosomal contents without killing a large fraction of the cells. The same effect may be obtained by using other photosensitizing compounds alone or associated with /linked to other molecules or particles used as vectors for directing the photosensitizers to endosomes/lysosomes or other intracellular compartments. Such vectors can be tissue or cell specific antibodies or other ligands that bind to the cell surface, thus increasing the uptake of the photosensitizer through receptor-mediated endocytosis. Another vector could be the use of reconstituted LDL-particles. These particles are also taken up by receptor-mediated endocytosis. The number of photosensitizer molecules per LDL particle and the binding to the LDL-particles can in this way be increased compared to prebinding to native LDL.

The present invention is not restricted to in vitro use, but may as well be used in vivo, either by in situ treatment or by ex vivo treatment followed by injection of the treated cells. The uptake into endosomes and lysosomes can be enhanced in the same manner as described above for in vitro treatment. All tissues can be treated as long as the photosensitizer is taken up by the target cells and the light can be properly delivered.

The present invention is based on both a photosensitizer and light. The light must be absorbed by the photosensitizer or indirectly induce an excited state of the photosensitizer. The wavelength region of use will therefore depend on the photosensitizer. The exposure light does not need to be monochromatic or collimated. Every light source emitting the appropriate wavelengths can be used.

Surprisingly the photodynamic action according to the present investigation seems to neutralize the potentially cytotoxic effect of releasing the lysosomal content. The present authors have thus established that lysosomal cathepsin is substantially inhibited by the fotodynamic action of $TPPS_{2a}$ in a culture of

NHIK 3025 cells. This was a surprising effect of the present invention and assists in maintaining the viability and functionality of the cells after transporting molecules into cytosol by disrupting endosomal/lysosomal membranes.

Examples of experimental and clinical utilization

1) Cancer treatment.

Several photosensitizers accumulate preferentially in neoplastic tissues, the selectivity for a tumor over the surrounding tissue being usually a factor of 2-3, but this factor may in some cases, such as for brain tissues, be higher, i.e. up to 30. Molecules which may be of clinical interest for treatment of cancer, but is restricted by a low or no uptake into the cytosol can be introduced into the cytosol by means of the present invention. Gelonin, as exemplified below, is an example of such a molecule. Several other molecules, either alone or linked to other molecules (e.g. antibodies, transferrin, photosensitizers, apoB on reconstituted LDL-particles) can be used. The advantage of such a combination treatment would be 1) enhanced cytotoxic effect in deeper layers of the tumor tissues since low and subtoxic doses of light are sufficient for disruption of lysosomes and endosomes; 2) enhanced specificity of the toxin since PDT is only given to the area of tumor localization.

2) Gene therapy

Gene therapy, i.e. therapeutic transfer of genes to the patients cells, is promising as a method for treating many genetic disorders such as cancer, cystic fibrosis, cardiovascular diseases and many other diseases. The main problem today is the transfection which must occur in vivo or in some cases can be performed ex vivo. Today, the most frequently used vector, i.e. the structure that helps delivering the DNA molecules into the cells, is different types of viruses, especially retro- and adenoviruses. The drawbacks of such methods are low stability of the vector, limited specificity, low yield and introduction of virus-DNA into human cells.

DNA, either as antisense DNA or whole genes, can be introduced into cells by the aid of photochemically induced disruption of endosomes and/or lysosomes. The treatment can be performed in vivo.

3) Experimental utilization

The present invention can be used to introduce a wide variety of molecules into cells in culture, e.g., genes, antibodies, manipulated proteins and compounds usually not permeable to the plasma membrane.

The present invention is further illustrated by the following non-limiting examples.

Example 1. This example demonstrates that photodynamic treatment releases a protein synthesis inhibiting compound into the cytosol.

A number of plant toxins kills cells by entering the cytosol and inactivating enzymatically the ribosomal function. The most cytotoxic plant proteins consist of 2 polypeptide chains, A and B, linked together by disulfide bridges. The function of chain B is to bind the protein to the surface of the cells, while chain A contains the enzymatic activity. Gelonin is a plant toxin which efficiently inhibits protein synthesis in cell-free systems, but has little or no effect on intact cells. The low cytotoxic effect on intact cells is probably due to the lack of a B chain in gelonin.

NHIK 3025 cells were incubated with $TPPS_{2a}$ (Formula I) and gelonin, separately or together for 18 h, followed by 1 h in $TPPS_{2a}$ and gelonin-free medium before the cells were exposed to light. Protein synthesis was measured 24 h after exposure to light. The photodynamic treatment, which kills 10-20% of the cells alone, reduced the protein synthesis by 30-40% (Fig.2). As seen in Fig.2 gelonin alone in the presence or absence of light inhibits protein synthesis to some extent. However, protein synthesis can be completely inhibited by combining PDT and gelonin with an IC_{50}=0.2 µg/ml gelonin. Thus in absence of the photodynamic treatment the gelonin essentially did not enter cytosol. This example indicates that $TPPS_{2a}$ and light can be used to introduce functionally intact macromolecules into the cellular cytosol.

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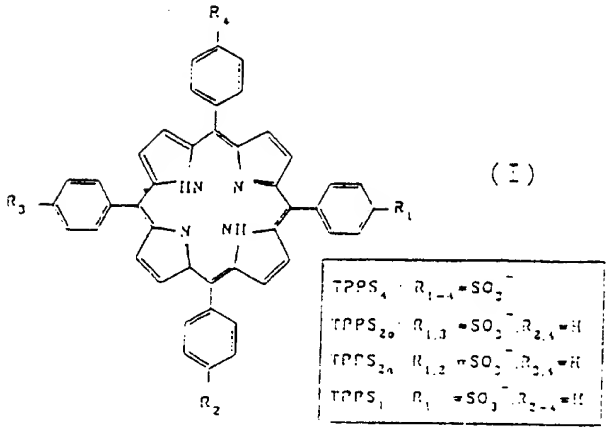

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Example 2. This example illustrates how the dose of light (with the wavelength which is absorbed by the dye) can be used to decide the size of the surviving cell fraction.

NHIK 3025 celles were incubated with $TPPS_{2a}$ and geloning according to the design of Example 1.

Clonogenic survivial of the cells was measured 24 h after exposure to light. As illustrated in Fig. 3 virtually all cells were killed with $TPPS_{2a}$ and light when the light exposure was increased. This is in accordance with prior art regarding killing unwanted cells with PDT. When gelonin is added the survival rate drops due to the inhibiting effect of gelonin on protein synthesis, showing that gelonin now is released in the cytosol. Increased concentration of added gelonin leads to more gelonin in the cytosol, as indicated by an increased sensitivity of the cells to photo inactivation.

The present invention thus offers the possibility to set a level of survival in each case and select a combination of photosensitizing compound and light exposure which will keep the wanted fraction of cells alive.

Example 3. This example illustrates how changing light doses control the amount of gelonin released in the cytosol, as determined by the relative protein synthesis.

NHIK 3025 cells were incubated with $TPPS_{2a}$ and gelonin according to the design of Example 1.

Figure 4 shows that light doses above the toxic dose of 50 sec increased the gelonin fraction in cytosol as determined by the relative protein synthesis.

Example 4-11 demonstrates use of the method according to the invention on different cell lines and with different photosensitizers and toxins. The intracellular location of the photosensitizers are lysosomal ($TPPS_4$, $TPPS_{2a}$, $AlPcS_{2a}$) and extralysosomal (3-THPP). The following abbrevations are used: $AlPcS_{2a}$ for aluminium phtalocyanine with 2 sulfonate groups on adjacent phenyl rings; $TPPS_4$ for meso-tetraphenylporphine with 4 sulfonate groups; TPPS2a for meso-tetraphenylporphine with 2 sulfonate groups on adjacent phenyl rings; 3-THPP for tetrahydroxylphenyl porphine. The used cell lines are carcinoma cells in situ from human cervix (NHIK 3025), Chinese hamster lung fibroblasts (V 79), SV40-transformed African Green monkey kidney (CV1-simian fibroblasts-like cells) (Cos-7) human osteosarcoma cells (OHS) and small cell lung cancer cells (H146). All experiments were designed as in Example 1.

Example 4. This example relates to use of the photosensitizer $AlPcS_{2a}$ in V79 cells with/without gelonin as the toxin (Fig. 5). By selecting a specific light dose (irradiation time) it is demonstrated that, without the toxin very little cell damage is produced as illustrated by the small reduction in protein synthesis, while with gelonin the protein synthesis is profoundly reduced. This shows the transport of gelonin molecules into cell cytoplasma via lysosomes without essentially damaging the cells even though the intracellular localization of $AlPcS_{2a}$ is lysosomal (Moan, J., Berg, K., Anholt, H. and Madslien, K. (1994). Sulfonated aluminium phtalocyanines as sensitizers for photochemotherapy. Effects of small light doses on localization, dye fluorescence and photosensitivity in V79 cells. Int. J. Cancer 58: 865-870).

Example 5. This example demonstrates transport of the toxin gelonin into H146 cells without essentially affecting the viability of the cells. (Fig. 6). $TPPS_{2a}$ is known to be lysosomal located in the cell. (Berg, K., Western, A., Bommer, J. and Moan, J.(1990) Intracellular localisation of sulfonated meso-tetraphenylporphines in a human carcinoma cell line. Photochem. Photobiol. 52:481-487; Berg, K., Madslien, K., Bommer J.C., Oftebro, R., Winkelman, J.C. and Moan, J. (1991). Light induced relocalization of sulfonated meso-tetraphenylporphines in NHIK 3025 cells and effects of dose fractionation. Photochem. Photobiol. 53:203-210; Berg, K and Moan, J. (1994) Lysosomes as photochemical targets. Int. J. Cancer. 59:814-822).

BNSDOCID: <WO 9607432A1 >

Example 6. This example demonstrates the method according to the invention in V79 cells using $TPPS_{2a}$ as photosensitizer (Fig. 7).

Example 7 This example demonstrates transport into NHIK 3025 cells of the toxin agrostin using the photosensitizer TPP_{2a} (Fig. 8).

Example 8 This example demonstrates transport of the toxin saporin into NHIK 3025 cells using TPP_{2a} (Fig. 9).

Example 9 This is a comparison example demonstrating that when a photosensitizer (3-THPP) which does not enter endocytic vesicles (i.e. endosomes and lysosomes) (Peng, Q., Danielsen, H.E. and Moan, J. (1994) Potent photosensitizers for photodynamic therapy of cancer: Applications of confocal laser scanning microscopy for fluorescence detection of photosensitizing fluorophores in neoplastic cells and tissues. In: Proceedings of Microscopy, Holography, and Interferometry in Biomedicine. SPIE Vol. 2083:71-82), there is no significant difference between the protein synthesis effect of 3THPP with or without gelonin (Fig. 10). Thus gelonin is not transported into the cytosol of the cells.

Example 10 This example demonstrates the transport of gelonin into COS-7 cells by using $TPPS_{2a}$ according to the invention (Fig. 11).

Example 11 This example demonstrates the transport of gelonin into OHS cells by using $TPPS_{2a}$ according to the invention. (Fig. 13). In this cell line there is a considerable protein degradation in the lysosomes, which in the present example is inhibited by incubating the cells for 4 hours in either 50 μM chloroquine or 10 mM NH_4Cl.

Example 12 Similar to example 1 this example demonstrates transport of gelonin into NHIK 3025 cells as a function of the gelonin concentration when the cells were incubated with $TPPS_4$ and different concentrations of gelonin, and exposed to light (Fig. 12). When the cells were incubated with gelonin alone and exposed to light, or incubated with TPPS4 and gelonin without exposure to light, no transport of gelonin into the cells was obtained.

The examples demonstrate that different molecules can be introduced into the cells cytosol in a wide variety of cells using different photosensitizers and doses

of light. Exogenous molecules can be introduced to the cellular cytosol after doses of photosensitizers and light which do not kill the cells, as long as the molecules to be introduced and the photosensitizers are transported to the same cellular compartments. The photochemical effect on a biological compartment is dependent upon the amount of photosensitizers in that compartment, the dose of light applied and the spectral properties of the light source. The best way to evaluate photochemical effects on cells in culture is therefore to measure cell survival 24 hours or more after treatment. There is a good correlation between the effect on cell killing and inhibition of protein synthesis 24 hours after treatments as presented above (data not shown).

CLAIMS

1. Method for introducing molecules into the cytosol of living cells by using photosensitizing compounds and light absorbed by the photosensitizing compound
characterized in that

a) the photosensitizing compound, optionally the vector molecules and the molecule(s) to be transported, which are bond together or separate, are applied to the living cells and;

b) are then endocytically or in other ways translocated into endosomes, lysosomes or other intracellular membrane-restricted compartments, whereby the cells are;

c) exposed to light with suitable wavelengths according to the absorption spectrum of the photosensitizing compound, in varying doses dependent upon the wanted survival rate such that the endosomal, lysosomal or other intracellular compartment membranes are disrupted and the molecules to be introduced in the cytosol are released in the cytosol, and that the light activated photosensitizing compound by itself does not kill the majority of the cells.

2. Method according to claim 1,
characterized in that the molecules which are released in the cytosol are selected from a group containing DNA, oligo(deoxy)nucleotides, mRNA, antisense DNA, sugars, proteins, peptides, membrane impermeable drugs, other membrane impermeable molecules, and covalently or non covalently bonded combinations of the above mentioned molecules.

3. Method according to claim 1,
characterized in that the molecules which are released in the cytosol is selected from a group of substances modifying the protein synthetic activity.

4. Method according to claim 2,
characterized in that the compound which is released in the cytosol is gelonin, saporin or agrostin or any combination of these.

5. Method according to claim 1,
c h a r a c t e r i z e d in that the used photosensitizing compounds is selected from the group containing porphyrins, phtalocyanines, purpurins, chlorins, benzoporphyrins, napthalocyanines, cationic dyes, tetracyclines and lysosomotropic weak bases and derivates thereof.

6. Method according to claim 5,
c h a r a c t e r i z e d in that the used photosensitizing compound is tetraphenyl porphine with 2 sulfonate groups on adjacent phenyl groups ($TPPS_{2a}$), meso-tetraphenyl porphine with 4 sulfonate groups ($TPPS_4$) or aluminium phtalocyanine with 2 sulphonate groups on adjacent phenyl rings ($AlPcS_{2a}$) or any combination of these.

7. Method according to claim 1,
c h a r a c t e r i z e d in that the used vector molecules are site-directing molecules or enhancers which facilitate the uptake of photosensitizing compound or the molecules which are to be released into the cytosol.

8. Method according to claim 1,
c h a r a c t e r i z e d in that the fraction of survival cells is regulated by selecting the light dose in relation to concentration of the photosensitizing compound.

9. Composition for the modification of neoplastic and other cellular processes in warm-blooded animals or cell cultures containing both a compound to be released into the cytosol and a photosensitizer and carriers,
c h a r a c t e r i z e d in that it comprises a member selected from the group consisting of
a) a carrier for facilitating the translocation into intracellular compartments, such as endosomes, lysosomes or other intracellular compartments, having attached thereto both the cellular process-modifying compound to be released in the cytosol and a photosensitizing compound, which upon light activation rupture intracellular compartment membranes,
b) several of said carriers wherein at least one carrier has attached thereto the compound to be released in cytosol and another similar or not similar carrier has attached thereto the photosensitizing compound and
c) a mixture of carriers separately together with the compound to be released in the cytosol and the photosensitizing compound.

10. Composition according to claim 9,
characterized in that the said compound to be released in the cytosol is an antineoplastic compound.

11. Composition according to claim 10,
characterized in that the antineoplastic compound is gelonin.

12. Composition according to claim 9,
characterized in that the compound to be released in the cytosol is selected from a group containing DNA, oligo(deoxy)nucleotides, mRNA, antisense DNA, sugars, proteins, peptides, membrane impermeable drugs, other membrane impermeable molecules, covalently or non-covalently bonded combinations of the above mentioned molecules.

13. Composition according to claim 9,
characterized in that the photosensitizing compound is selected from a group containing porphyrins, phtalocyanines, purpurins, chlorins, benzoporphyrins, napthalocyanines, cationic dyes, tetracyclines and lysosomotropic weak bases and derivates thereof.

14. Composition according to claim 11,
characterized in that the photosensitizing compound is tetraphenylporphine with two sulfonate groups on adjacent phenyl groups.

15. Composition according to claim 9,
characterized in that the carriers are selected from site-directing molecyls or enhancers with facilitate the uptake of photosensitizing compounds or the molecyls which are to be released into the cytosol.

16. Kit for performing the method according to claim 1,
characterized in that it contains photosensitizing compounds and other compounds modifying the cell membrane transport, facilitating certain cell populations or other related functions.

FIG. 1

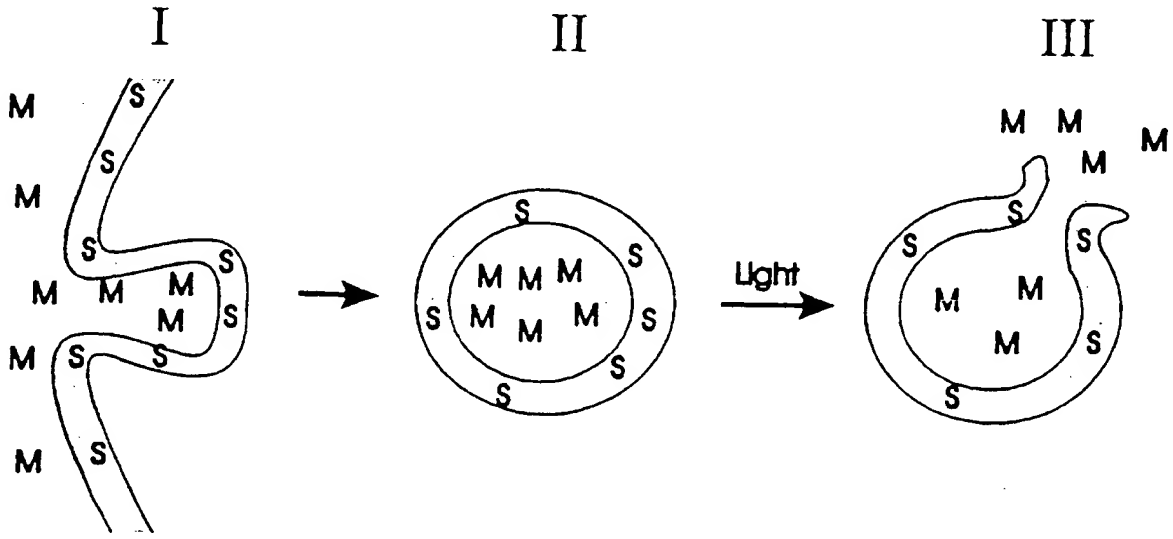
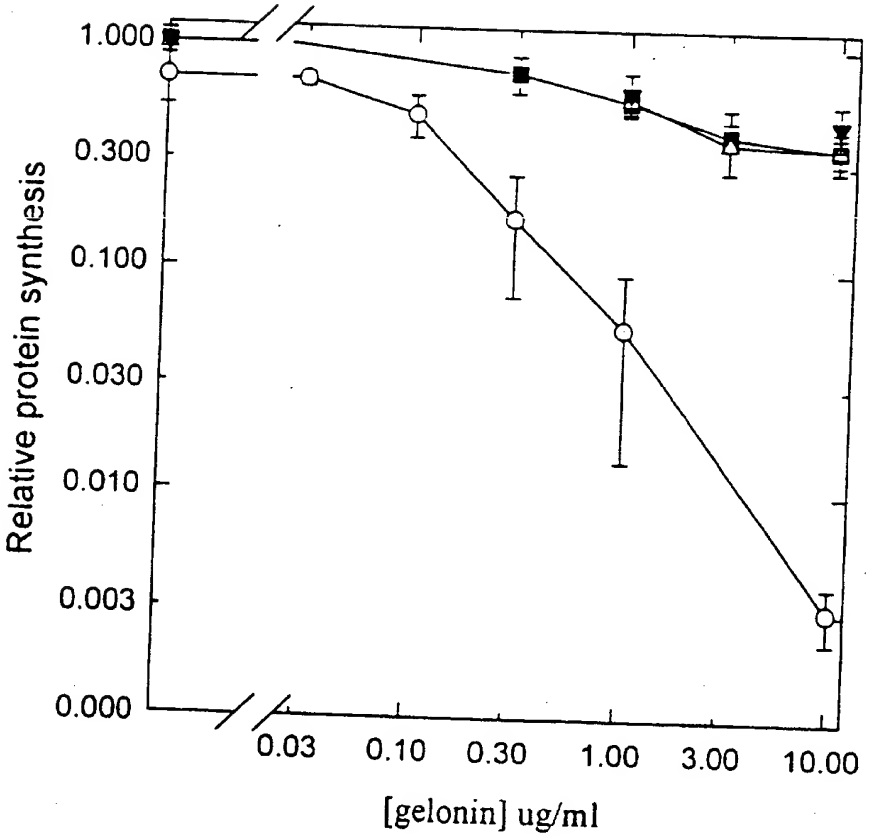


FIG. 2



BNSDOCID: <WO 9607432A1 I >

FIG. 3

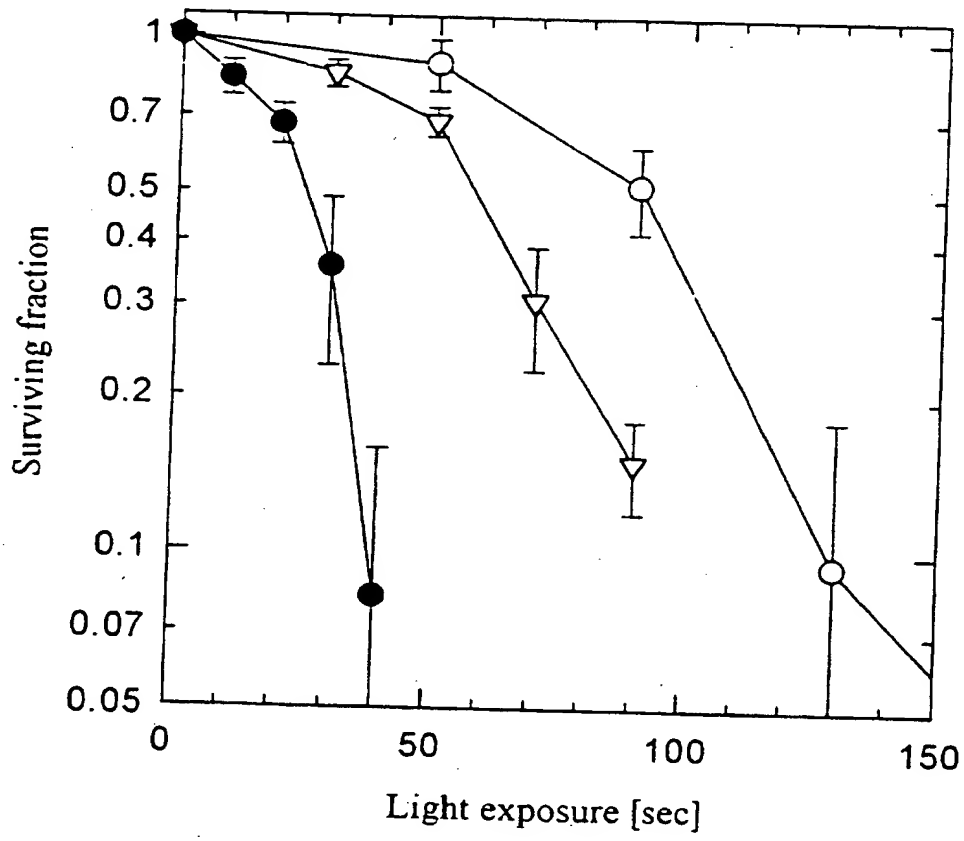
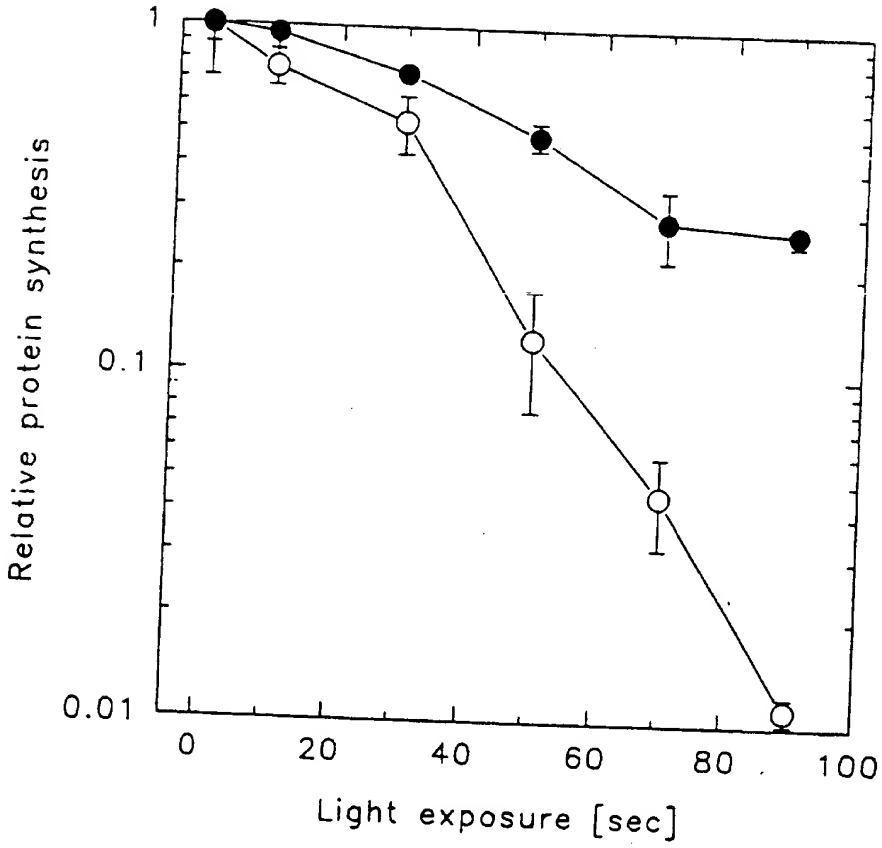

Surviving fraction
1
0.7
0.5
0.4
0.3
0.2
0.1
0.07
0.05
0
50
100
150
Light exposure [sec]

FIG. 4



BNSDOCID: <WO___9607432A1_I_>

FIG. 5

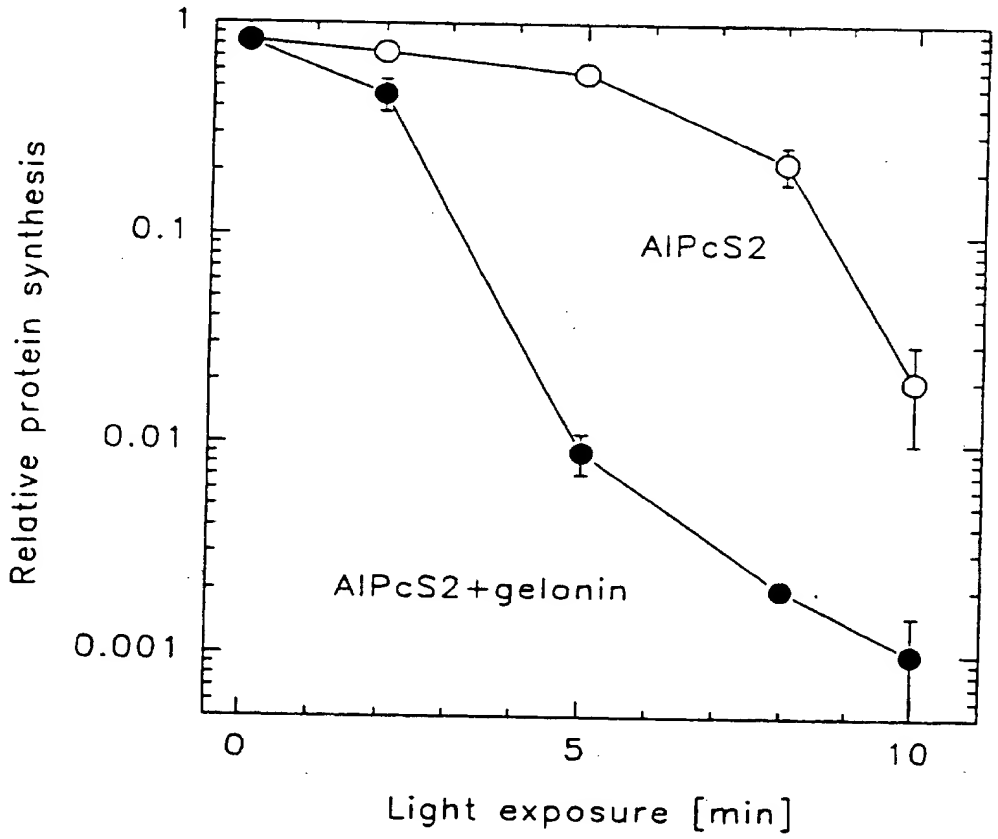
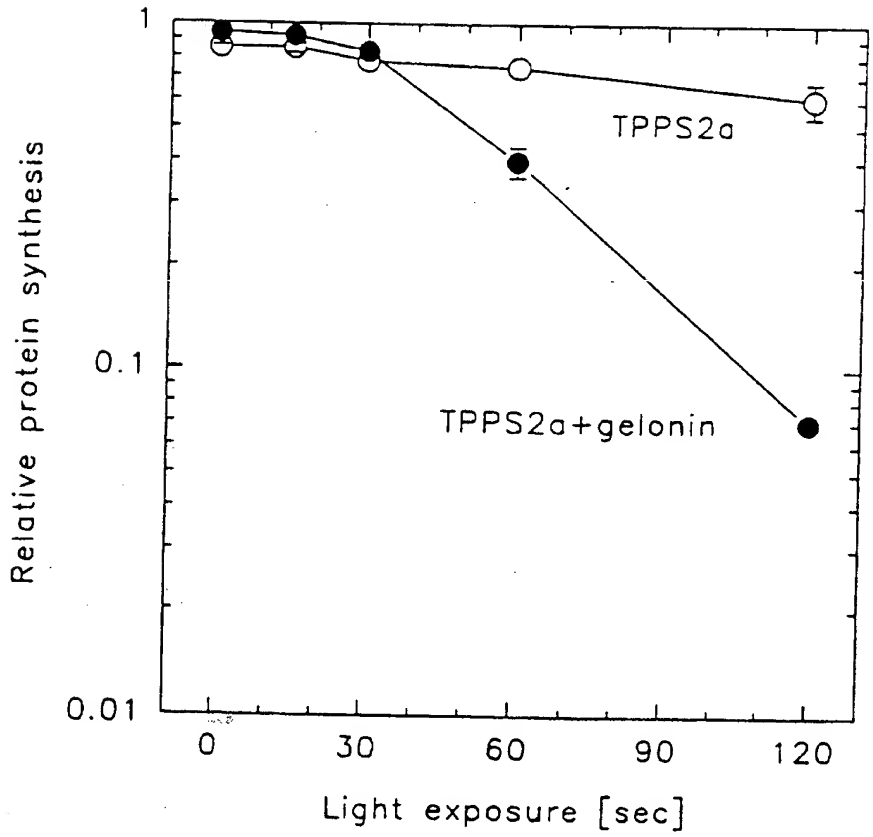

1
0.1
0.01
0.001
Relative protein synthesis
AlPcS2
AlPcS2+gelonin
0
5
10
Light exposure [min]

FIG. 6



BNSDOCID: <WO 9607432A1 I >

FIG. 7

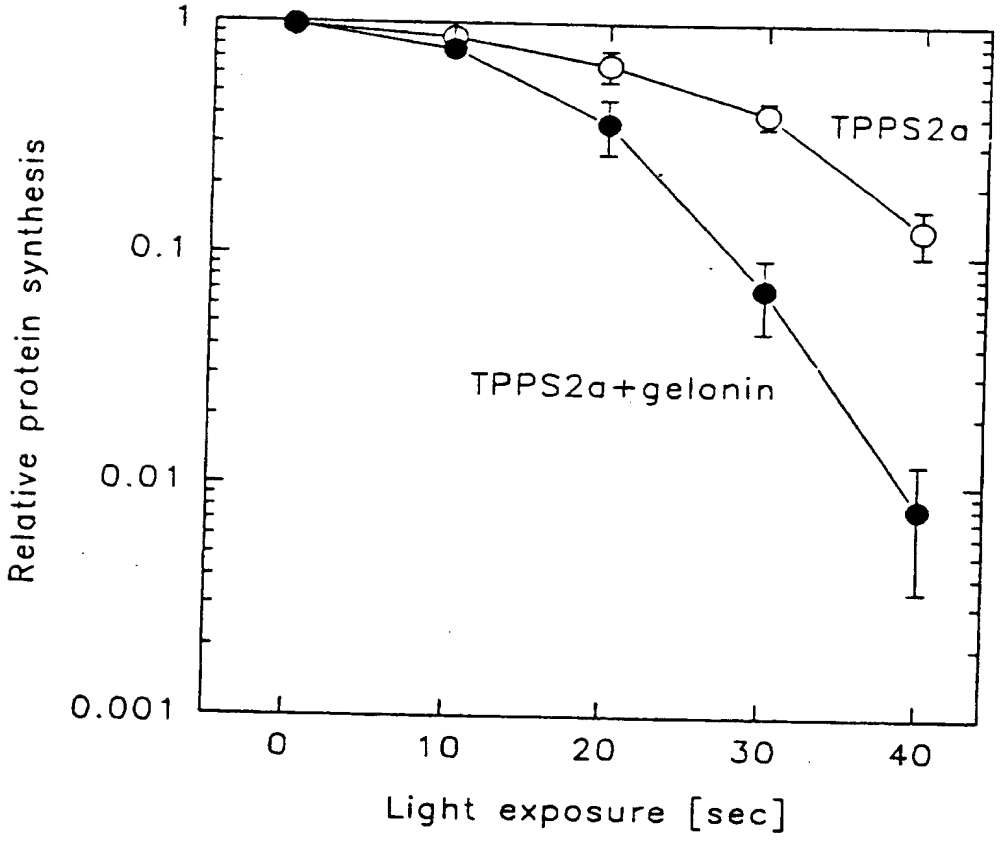
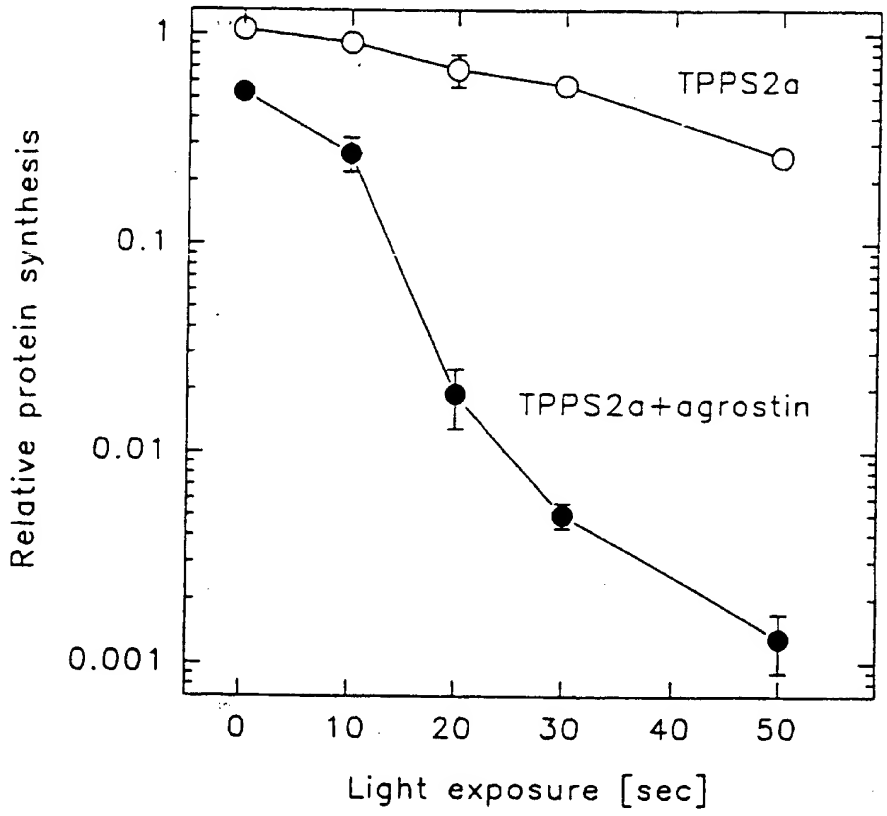

1
0.1
0.01
0.001
Relative protein synthesis
TPPS2a
TPPS2a+gelonin
0
10
20
30
40
Light exposure [sec]

FIG. 8



BNSDOCID: <WO 9607432A1 >

FIG. 9

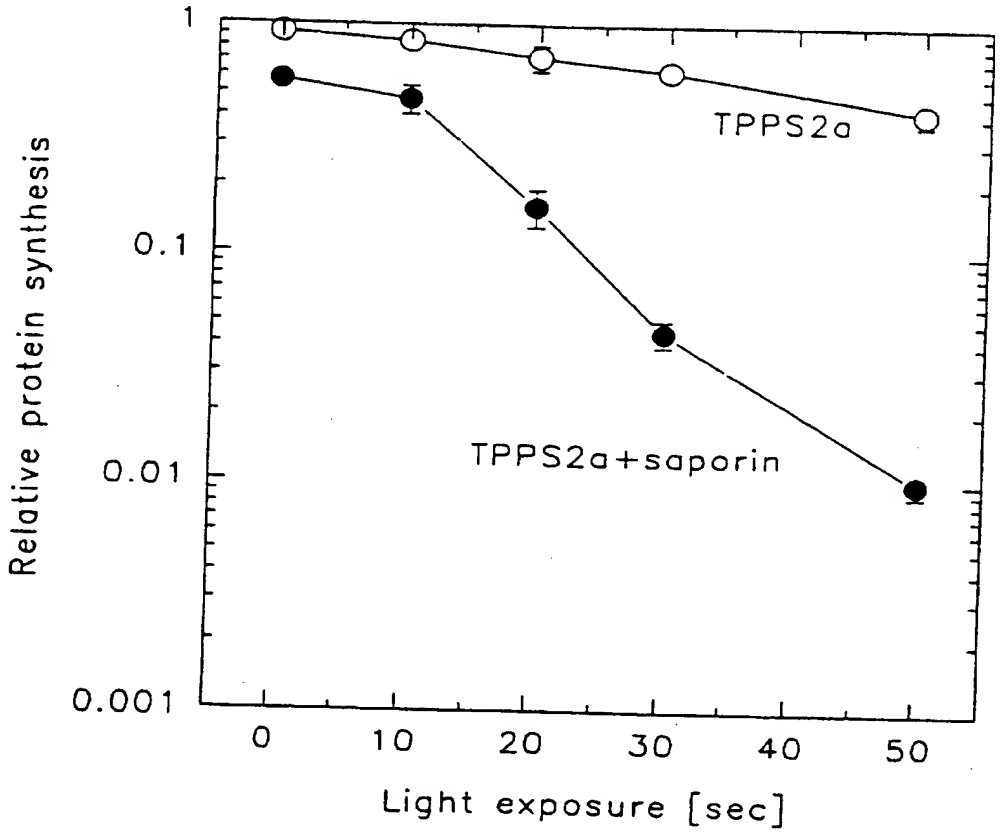
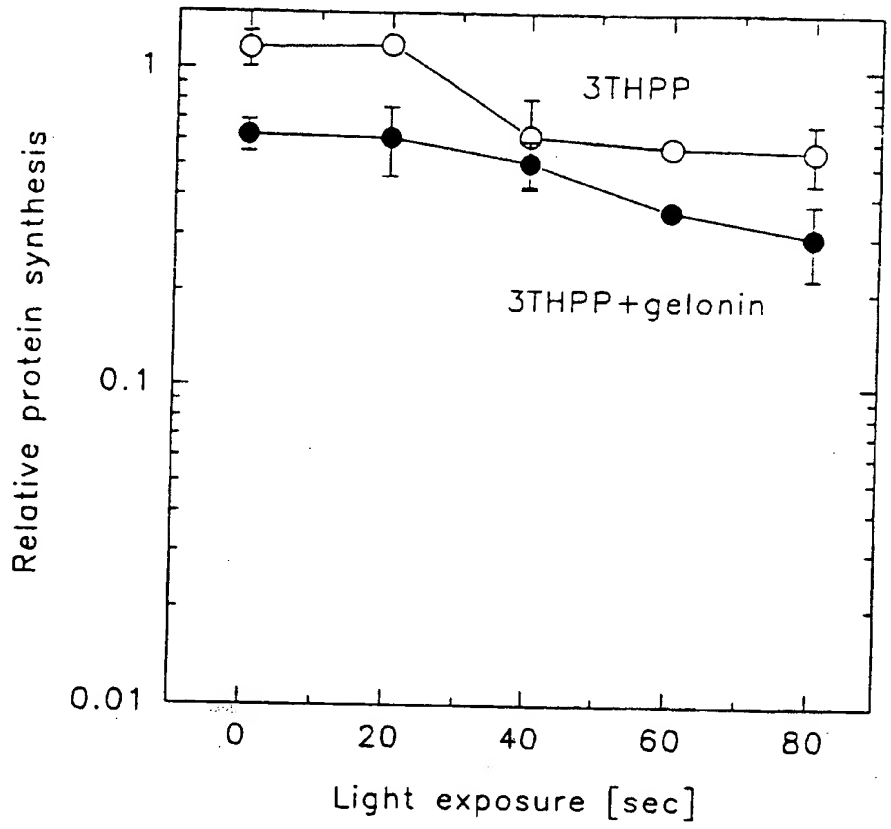

1
0.1
0.01
0.001
Relative protein synthesis
TPPS2a
TPPS2a+saporin
0
10
20
30
40
50
Light exposure [sec]

FIG. 10



BNSDOCID: <WO 9607432A1 >

FIG. 11

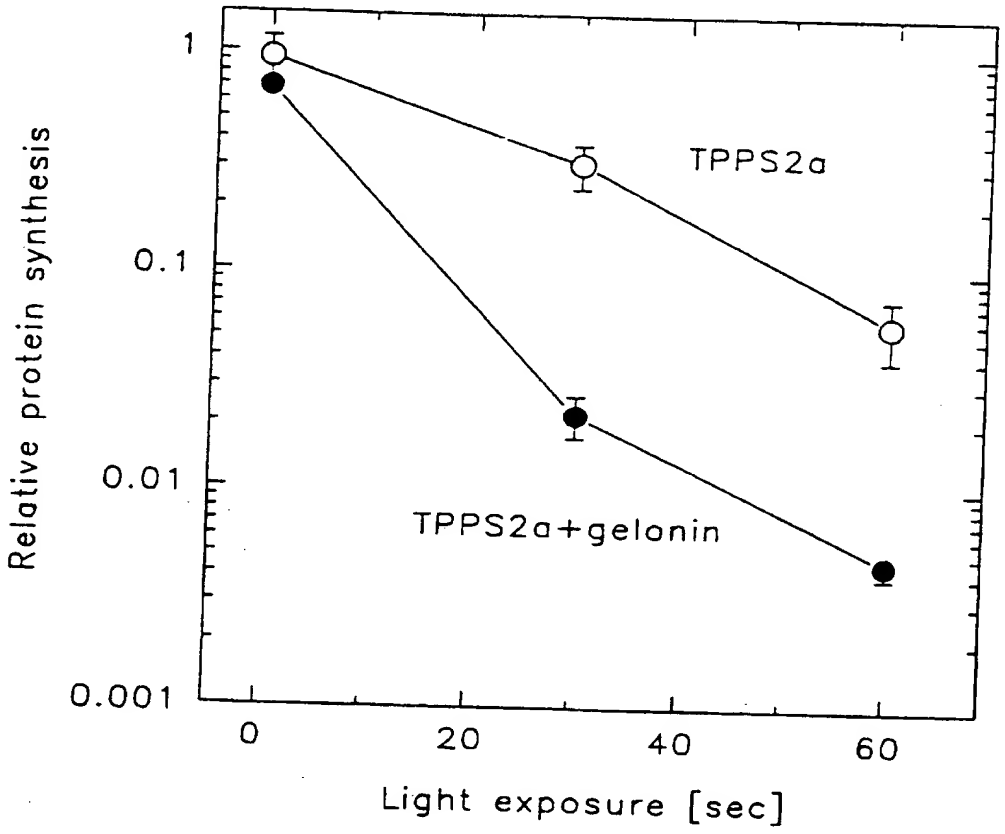
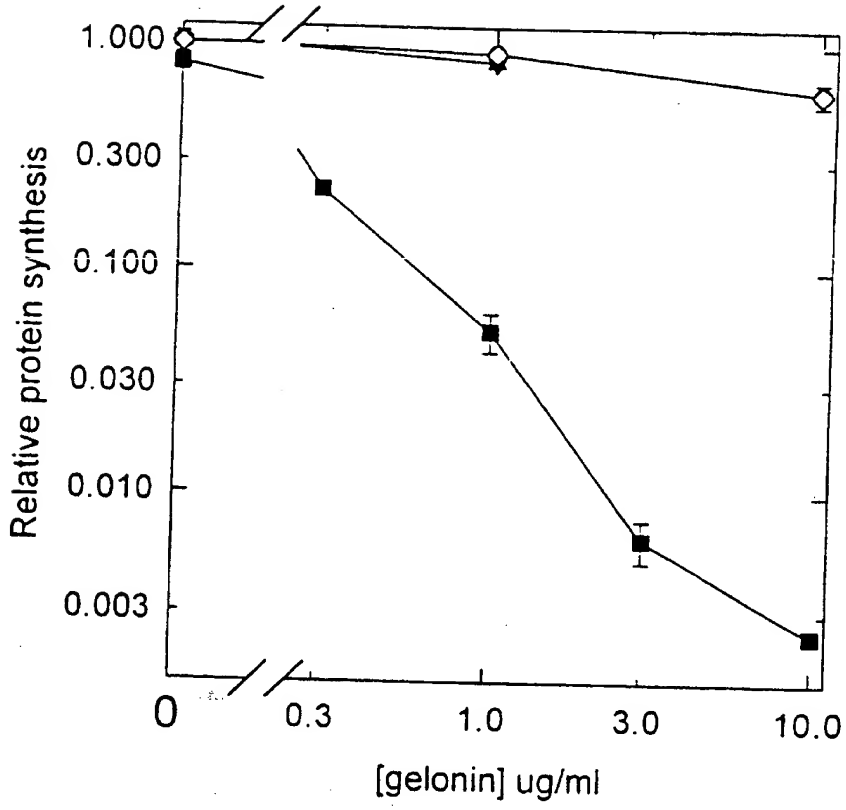
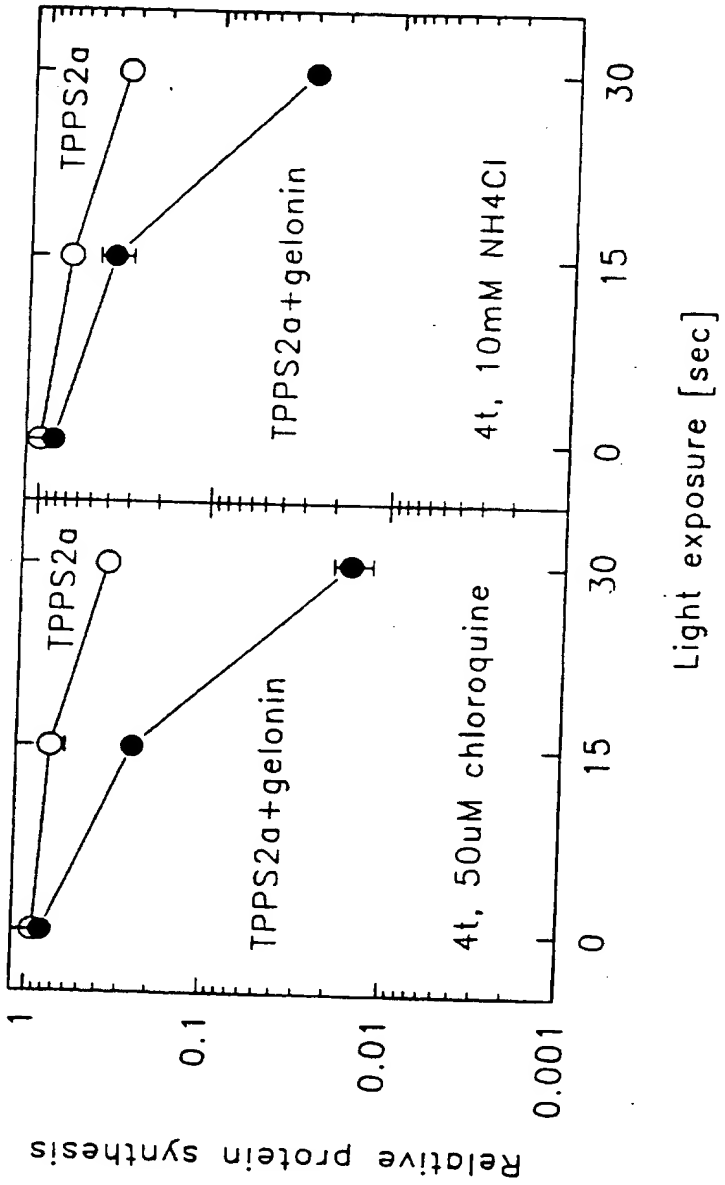

1
0.1
0.01
0.001
Relative protein synthesis
TPPS2a
TPPS2a+gelonin
0
20
40
60
Light exposure [sec]

FIG. 12



BNSDOCID: <WO 9607432A1 >

FIG. 13


Relative protein synthesis
1
0.1
0.01
0.001
TPPS2a
TPPS2a+gelonin
4t, 50uM chloroquine
TPPS2a
TPPS2a+gelonin
4t, 10mM NH4Cl
0
15
30
0
15
30
Light exposure [sec]

INTERNATIONAL SEARCH REPORT

International application No.
PCT/NO 95/00149

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: A61K 47/48, A61K 47/00
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

MEDLINE, BIOSIS, EMBASE, WPI, WPIL, US PATENT FULLTEXT DATABASES, SCISEARCH

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	GB 2209468 A (THE UNIVERSITY OF SALFORD), 17 May 1989 (17.05.89), page 6, line 10 - line 27; page 7, line 30 - line 31; page 9, line 16, the claims -- --------	1-16

☐ Further documents are listed in the continuation of Box C. ☒ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" erlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
21 December 1995	29 -12- 1995
Name and mailing address of the ISA/ Swedish Patent Office Box 5055, S-102 42 STOCKHOLM Facsimile No. +46 8 666 02 86	Authorized officer Carolina Palmcrantz Telephone No. +46 8 782 25 00

Form PCT/ISA/210 (second sheet) (July 1992)

BNSDOCID: <WO 9607432A1 >

INTERNATIONAL SEARCH REPORT

International application No.
PCT/NO 95/00149

Box I **Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.: 1-8
because they relate to subject matter not required to be searched by this Authority, namely:
Claims 1-8 relate to a method for treatment of the human or animal body by therapy since the claims are not restricted to a method performed in vitro (c.f. PCT Rule 39.1(iv)).
Nevertheless, a search has been executed for these claims.

2. [X] Claims Nos.: 16
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
see extra sheet

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II **Observations where unity of invention is lacking (Continuation of item 2 of first sheet)**

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.

2. [ ] As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.

PNSDOCID: WO 9607439A1 ( )

Claim 16 is not considered to be clear and concise with regard to the wording

"...and other compounds modifying the cell membrane transport, facilitating certain cell populations or other related functions". It is not clear which the intended "other modifying compounds" are (c.f. PCT Article 6).

BNSDOCID: <WO 9607432A1 >

INTERNATIONAL SEARCH REPORT

Information on patent family members

11/12/95

International application No.
PCT/NO 95/00149

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
GB-A- 2209468	17/05/89	NONE	